

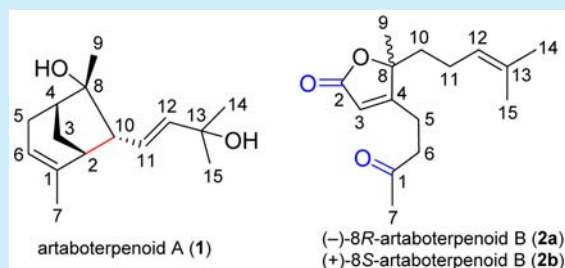
Artaboterpenoids A and B, Bisabolene-Derived Sesquiterpenoids from *Artabotrys hexapetalus*

Feng-Min Xi, Shuang-Gang Ma, Yun-Bao Liu, Li Li, and Shi-Shan Yu\*

State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China

## Supporting Information

**ABSTRACT:** Artaboterpenoids A and B (**1** and **2**), two novel bisabolene-derived sesquiterpenoids, were isolated from the roots of *Artabotrys hexapetalus*. Their structures with absolute configurations were elucidated by spectroscopic methods, and electronic circular dichroism (ECD) analyses. Notably, **1** featured a novel carbon skeleton with a new C-2–C-10 linkage, and **2a** and **2b**, a pair of enantiomers, represented the first examples of 1,2-seco-bisabolene-type sesquiterpene lactones. **2a** exhibited cytotoxic effects against HCT-116, HepG2, A2780, NCI-H1650, and BGC-823 cell lines with IC<sub>50</sub> values of 1.38–8.19  $\mu$ M. Plausible biogenetic pathways for artaboterpenoids A and B were proposed.



Bisabolene-type sesquiterpenoids are a very important family of natural products with structural diversity and various bioactivities, such as juvenile hormone,<sup>1</sup> antitumor,<sup>2</sup> and antimalarial<sup>3</sup> activities. Bisabolene was considered to be the biogenetic precursor of many sesquiterpenes.<sup>4</sup> For instance, amorphadiene, acoradiene, and cuprenene were derived from bisabolene by cyclization,<sup>5</sup> which was demonstrated by Tantillo with quantum chemical calculations.<sup>6</sup>

*Artabotrys hexapetalus* (L.f.) Bhandari was a Chinese folk medicine used for the treatment of malaria. Yingzhaosus A–D,<sup>3,7</sup> as representative bisabolene-type sesquiterpenoids, were discovered from the root of *A. hexapetalus*. In our continuing endeavor to discover structurally diverse and biologically interesting metabolites from traditional Chinese medicines, two bisabolene-derived sesquiterpenoids, artaboterpenoids A and B (**1** and **2**), were isolated from the root of *A. hexapetalus*. Artaboterpenoid A (**1**) possesses an unusual carbon skeleton formed by the new C-2–C-10 linkage (**1**), and (±)-artaboterpenoid B (**2a** and **2b**), a pair of enantiomers, were first reported as 1,2-seco-bisabolene-type sesquiterpene lactones. Herein, we report their isolation, structural elucidation, and cytotoxicity, as well as plausible biogenetic pathways.

Artaboterpenoid A (**1**) was isolated as a colorless oil with the molecular formula C<sub>15</sub>H<sub>24</sub>O<sub>2</sub>, as determined by (+)-HRESIMS data (*m/z* 259.1670 [M + Na]<sup>+</sup>, calcd for 259.1669), corresponding to four degrees of unsaturation. The <sup>13</sup>C and HSQC NMR data displayed three quaternary carbons (one olefinic and two oxygenated), six methines (three olefinic), two methylenes, and four methyls (Table 1). The <sup>1</sup>H NMR spectrum showed resonances attributable to a pair of *trans* olefinic protons [ $\delta_{\text{H}}$  6.23 (dd, *J* = 15.7, 9.9 Hz), 6.04 (d, *J* = 15.7 Hz)], one olefinic proton singlet ( $\delta_{\text{H}}$  5.33), and four tertiary methyls ( $\delta_{\text{H}}$  1.72, 1.51  $\times$  2, and 1.44) (Table 1). These data combined with the degrees of unsaturation suggested a two-ring system in the structure of **1**.

Table 1. <sup>1</sup>H NMR (500 MHz) and <sup>13</sup>C NMR (125 MHz) Data of Artaboterpenoids A and B

no.	artaboterpenoid A <sup>a</sup>		artaboterpenoid B <sup>b</sup>	
	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (J, Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (J, Hz)
1	139.2		205.7	
2	47.6	2.17, br s	171.8	
3	34.5	a 1.79, m b 1.75, m	114.5	5.65, t (1.8)
4	46.6	2.16, br s	174.9	
5	30.1	a 2.82, m b 2.16, m	20.8	2.48, m, 2H
6	120.4	5.33, br s <sup>c</sup>	39.8	2.82, m, 2H
7	25.3	1.72, s	29.8	2.23, s
8	79.5		89.2	
9	32.9	1.44, s	23.8	1.45, s
10	60.8	2.52, dd (9.9, 5.8)	36.9	a 1.89, m b 1.66, m
11	126.7	6.23, dd (15.7, 9.9)	21.6	a 1.96, m b 1.79, m
12	141.6	6.04, d (15.7)	122.5	5.02, m
13	70.2		132.4	
14	31.2	1.51, s	17.5	1.56, br s
15	31.0	1.51, s	25.4	1.66, br s
8-OH		4.92, br s		
13-OH		5.86, br s		

<sup>a</sup>Data were recorded in pyridine-*d*<sub>5</sub>. <sup>b</sup>Data were recorded in CDCl<sub>3</sub>.

<sup>c</sup>The peak width at half-height was 7.8 Hz.

Received: May 26, 2016

Published: June 24, 2016

Detailed analyses of the  $^1\text{H}$ – $^1\text{H}$  COSY and HSQC spectra were not able to lead to the assignment of the spin systems of  $\text{C}(10)\text{H}$ – $\text{C}(2)\text{H}$ – $\text{C}(3)\text{H}_2$ – $\text{C}(4)\text{H}$ – $\text{C}(5)\text{H}_2$  because of the three overlapped signals at 2.17 ppm (H-2, 4, and 5b) (Figure 1). Afterward, a six-membered ring A was established by the

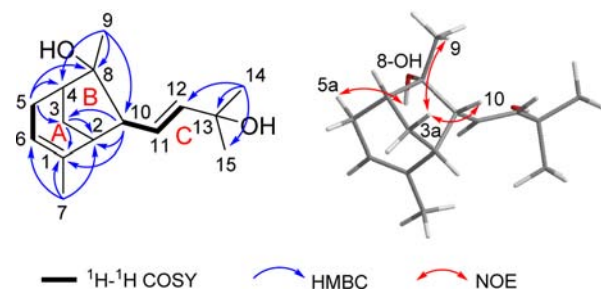


Figure 1. Key  $^1\text{H}$ – $^1\text{H}$  COSY, HMBC, and NOE correlations for **1**.

HMBC cross-peaks from  $\text{H}_3$ -7 to C-1, C-2, and C-6, from H-3b to C-1, C-2, and C-10, and from H-5a to C-3, C-4, and C-8, along with the  $^1\text{H}$ – $^1\text{H}$  COSY cross-peak between H-5a and H-6. Additionally, the HMBC cross-peaks from  $\text{H}_3$ -9 to C-4, C-8, and C-10 and from H-10 to C-1, C-2, and C-3 implied the presence of a five-membered ring B. Also, the side chain of ring B (fragment C) was deduced by the HMBC cross-peaks from  $\text{H}_3$ -14 to C-12, C-13, and C-15, along with the  $^1\text{H}$ – $^1\text{H}$  COSY cross-peaks of H-10/H-11/H-12. Thus, the planar structure of **1** was established as depicted (Figure 1).

The relative configuration of **1** was deduced from the interpretation of the nuclear overhauser effect (NOE) experiments. The remarkable NOE correlations from  $\text{H}_3$ -9 ( $\delta_{\text{H}}$  1.44) to H-3a ( $\delta_{\text{H}}$  1.79) and from H-10 ( $\delta_{\text{H}}$  2.52) to H-3a placed  $\text{H}_3$ -9, H-3a, and H-10 on the same face, while the strong NOE correlations from H-5a ( $\delta_{\text{H}}$  2.82) to C-8-OH ( $\delta_{\text{H}}$  4.92) placed these protons on the other side (Figure 1). Therefore, there were only two possible structures for **1**, with absolute configurations of **1A** (2*S*,4*R*,8*R*,10*S*) and **1B** (2*R*,4*S*,8*S*,10*R*) (Figure 2).

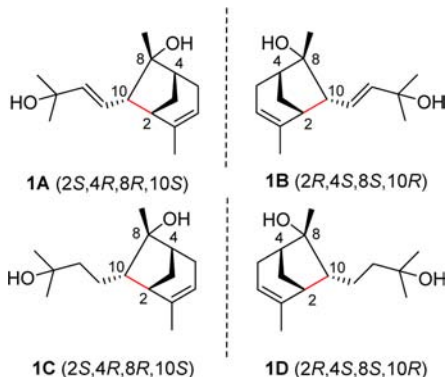


Figure 2. Structures of model compounds **1A** and **1C**, and **1B** and **1D** in the ECD calculations.

The absolute configuration of **1** was established by experimental and theoretical ECD after initially unsuccessful attempts to obtain single crystals of **1** and its *p*-bromobenzoate derivative. The ECD spectrum of **1** showed an intense negative Cotton effect at 208 nm ( $\Delta\epsilon$  –18.04), which predominately resulted from the  $\pi_x \rightarrow \pi_x^*$  electronic transition of the  $\Delta^{1(6)}$  double bond of the cyclohexene ring,<sup>8</sup> rather than the  $\Delta^{11}$  double bond. Time-dependent density functional theory<sup>9</sup> at the B3LYP/

6-31G(d) level was employed to demonstrate the aforementioned deduction by comparing the calculated ECD spectra of **1A** and **1C**, and of **1B** and **1D** (Figure 2). The Cotton effects of **1A** and **1C**, and of **1B** and **1D** were the same in the positions and signs but different in the relative intensities, respectively (Figure 3). Thus, the absolute configuration of **1** was determined by

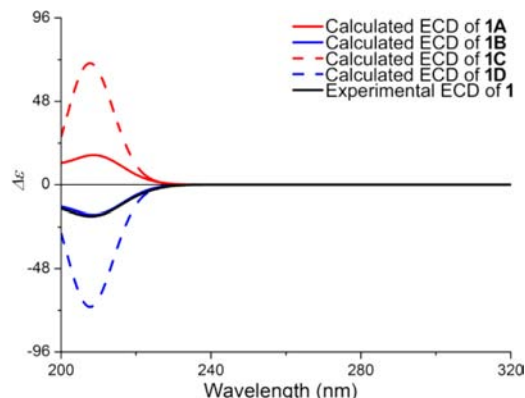


Figure 3. Experimental ECD spectrum of **1** and calculated ECD spectra of **1A** and **1C**, and **1B** and **1D**.

comparison of the experimental and calculated ECD spectra. The experimental ECD spectrum of **1** showed excellent agreement with the calculated spectrum of **1B** (2*R*,4*S*,8*S*,10*R*) (Figure 3).

Artaboterpenoid **2** possessed a molecular formula of  $\text{C}_{15}\text{H}_{23}\text{O}_3$  as established by (+)-HRESIMS data ( $m/z$  251.1640 [ $\text{M} + \text{H}$ ]<sup>+</sup>, calcd for 251.1642). The IR spectrum exhibited the presence of ester and ketone carboxyl groups (1751, 1720  $\text{cm}^{-1}$ ) and a double bond (1639  $\text{cm}^{-1}$ ). The analysis of its  $^1\text{H}$ ,  $^{13}\text{C}$ , and HSQC NMR data revealed the presence of five quaternary carbons [one ketone carboxyl ( $\delta_{\text{C}}$  205.7), one ester carboxyl ( $\delta_{\text{C}}$  171.8), two olefinic carbons ( $\delta_{\text{C}}$  174.9 and 132.4), and one oxygenated ( $\delta_{\text{C}}$  89.2)], two  $\text{sp}^2$  methines [ $\delta_{\text{H}}$  5.65 (t, 1.8 Hz) and 5.02 (m);  $\delta_{\text{C}}$  122.5 and 114.5), four  $\text{sp}^3$  methylenes ( $\delta_{\text{C}}$  39.8, 36.9, 21.6, and 20.8), and four tertiary methyls ( $\delta_{\text{H}}$  2.23, 1.66, 1.56, and 1.45;  $\delta_{\text{C}}$  29.8, 25.4, 23.8, and 17.5) (Table 1). These characteristic signals, with the five degrees of unsaturation, suggested that **2** was a sesquiterpene lactone with a single-ring system in the structure.

The planar structure of **2** was constructed by detailed analyses of its  $^1\text{H}$ – $^1\text{H}$  COSY and HMBC spectra. The  $^1\text{H}$ – $^1\text{H}$  COSY correlations from H-5 to H-6 as well as the HMBC correlations from  $\text{H}_3$ -7 to C-1 and C-6 established the connectivity of C-7–C-1–C-6–C-5. The  $^1\text{H}$ – $^1\text{H}$  COSY correlations from H-11 to H-10 and H-12, along with HMBC correlations from  $\text{H}_3$ -14 to C-12, C-13, and C-15, indicated the linkage of C-10–C-11–C-12–C-13(–C-15)–C-14. The HMBC correlations from H-3 to C-2, C-4, and C-8 and from  $\text{H}_3$ -9 to C-4 and C-8 implied the presence of an  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone unit (Figure 4). Furthermore, the HMBC correlations from  $\text{H}_2$ -5 to C-3, C-4, and C-8 and from  $\text{H}_2$ -6 to C-4 unambiguously confirmed the connectivity of C-4 and C-5, whereas the HMBC correlations from  $\text{H}_2$ -10 to C-8 and from  $\text{H}_2$ -11 to C-8 established the linkage of C-10 and C-8 (Figure 4). Thus, the planar structure of **2** is shown in Figure 4. It is particularly noteworthy that **2** is the first example of a 1,2-secobisabolene-type sesquiterpene lactone.

ECD has proven to be a powerful and reliable method for determining the absolute configuration of lactones.<sup>10</sup> However, Cotton effect resulting from  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone was not observed in the ECD spectrum of **2** (Figure S24, Supporting

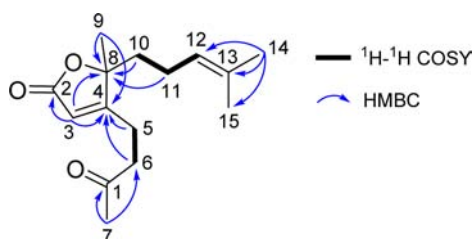


Figure 4. Key  $^1\text{H}$ – $^1\text{H}$  COSY and HMBC correlations for **2**.

Information). Furthermore, subsequent HPLC separation on a Chiralcel OZ-H column (Daicel Chemical Industries, Ltd., Tokyo, Japan) was successful in resolving two stereoisomers (**2a** and **2b**) (Figure 5), which were opposite in terms of the ECD

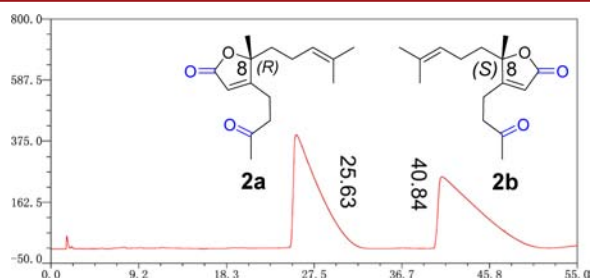


Figure 5. HPLC separation chromatogram of **2** on a Chiralcel OZ-H column.

curves (Figures S25 and S26, Supporting Information) and specific rotations  $\{[\alpha]_{\text{D}}^{20} -42.1$  ( $c$  0.1, MeOH) for **2a** and  $[\alpha]_{\text{D}}^{20} +43.6$  ( $c$  0.1, MeOH) for **2b**} but identical in terms of HRESIMS and NMR data of **2** (Figures S12–S20, Supporting Information). The observed Cotton effect around 220 nm due to the  $\pi \rightarrow \pi^*$  transition of the  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone chromophore was correlated directly to the chirality of the  $\gamma$ -carbon of butenolide. According to the helix rule,<sup>11</sup> a negative Cotton effect at 223.5 nm in the ECD spectrum of **2a** revealed that the C-10–C-8–C-4=C3 system possessed a left-handed helicity (counterclockwise) and suggested an 8*R* absolute configuration for **2a**, whereas a positive Cotton effect at 222.5 nm in the ECD spectrum of **2b** corresponded to a right-handed helicity (clockwise), indicating an 8*S* absolute configuration for **2b** (Figure 6). Therefore, **2a** and **2b** were named (–)-8*R*- and (+)-8*S*-artaboterpenoid B, respectively.

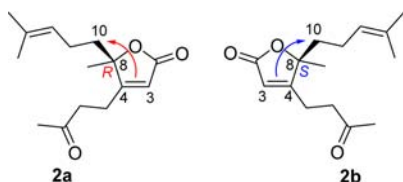
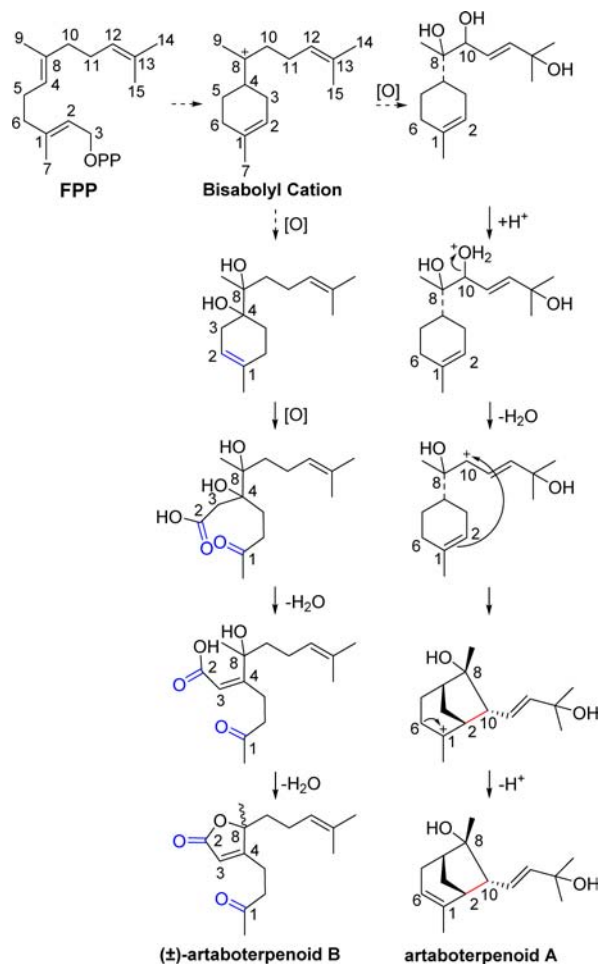


Figure 6. Helix rules for **2a** and **2b**.

Compounds **1**, **2a**, and **2b** were evaluated for their cytotoxicity against five human cancer cell lines (HCT-116, HepG2, BGC-823, NCI-H1650, and A2780) using the MTT method reported previously.<sup>12</sup> Taxol was used as a positive control ( $\text{IC}_{50}$  values of 31.6, 17.8, 1.17, 69.5, and 33.0 nM, respectively). Compound **2a** exhibited cytotoxicity against all five human tumor cell lines tested, with  $\text{IC}_{50}$  values of 1.38, 3.30, 6.51, 8.19, and 2.14  $\mu\text{M}$ , respectively, and the others were inactive ( $\text{IC}_{50} > 10 \mu\text{M}$ ).

Biosynthetically, artaboterpenoids A and B could be generated from bisabolene, as shown in Scheme 1. The route of

#### Scheme 1. Plausible Biosynthetic Pathways of Artaboterpenoids A and B



artaboterpenoid A is primarily involved in oxidation and cyclization, while the pathway of (±)-artaboterpenoid B is principally based on a ring-opening reaction by double bond oxidation and lactonization by dehydration.

In summary, we discovered two sesquiterpenoids from the roots of *A. hexapetalus*, including a novel endocyclic sesquiterpene with an unusual skeleton by the new C-2–C-10 linkage (**1**) and an unprecedented 1,2-secocyclic sesquiterpene lactone (**2**). By chiral chromatography, we achieved the chiral resolutions of a pair of enantiomers, (±)-artaboterpenoid B (**2a** and **2b**). The potent cytotoxicity and structural simplicity of **2a** make it a promising lead for the development of antitumor agents. We provide these examples of sesquiterpenes at the biosynthetic crossroad for bisabolane-type sesquiterpenoids.

#### ■ ASSOCIATED CONTENT

##### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.6b01519.

Detailed experimental procedures, 1D and 2D NMR, MS, IR, UV, and ECD spectra (PDF)

## AUTHOR INFORMATION

### Corresponding Author

\*E-mail: [yushishan@imm.ac.cn](mailto:yushishan@imm.ac.cn).

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

This work was supported by grants from the National Natural Science Foundation of China (No. 21132009) and the National Science and Technology Project of China (No. 2012ZX09301002-002). The authors are grateful to the Department of Instrumental Analysis at our institute for the spectroscopic measurements.

## REFERENCES

- (1) Slama, K.; Williams, C. M. *Proc. Natl. Acad. Sci. U. S. A.* **1965**, *54*, 411.
- (2) Kupchan, S. M.; La Voie, E. J.; Branfman, A. R.; Fei, B. Y.; Bright, W. M.; Bryan, R. F. *J. Am. Chem. Soc.* **1977**, *99*, 3199–3201.
- (3) Liang, X. T.; Yu, D. Q.; Wu, W. L.; Deng, H. C. *Acta Chim. Sinica* **1979**, *37*, 215–230.
- (4) Parker, W.; Roberts, J. S.; Ramage, R. Q. *Rev., Chem. Soc.* **1967**, *21*, 331–363.
- (5) Rücker, G. *Angew. Chem., Int. Ed. Engl.* **1973**, *12*, 793–806.
- (6) (a) Hong, Y. J.; Tantillo, D. J. *J. Am. Chem. Soc.* **2009**, *131*, 7999–8015. (b) Hong, Y. J.; Tantillo, D. J. *J. Am. Chem. Soc.* **2014**, *136*, 2450–2463.
- (7) (a) Liang, X. T.; Yu, D. Q.; Pan, W. D. *Acta Chim. Sinica* **1979**, *37*, 231–240. (b) Zhang, L.; Zhou, W. S.; Xu, X. X. *J. Chem. Soc., Chem. Commun.* **1988**, *8*, 523–524.
- (8) (a) Scott, A. I.; Wrixon, A. D. *Tetrahedron* **1970**, *26*, 3695–3715. (b) Scott, A. I.; Wrixon, A. D. *Tetrahedron* **1971**, *27*, 4787–4819.
- (9) Berova, N.; Bari, L. D.; Pescitelli, G. *Chem. Soc. Rev.* **2007**, *36*, 914–931.
- (10) (a) Beecham, A. F. *Tetrahedron* **1972**, *28*, 5543–5554. (b) Uchida, I.; Kuriyama, K. *Tetrahedron Lett.* **1974**, *15* (43), 3761–3764.
- (11) (a) Gawronski, J. K.; van Oeveren, A.; van der Deen, H.; Leung, C. W.; Feringa, B. L. *J. Org. Chem.* **1996**, *61*, 1513–1515. (b) Liu, Y.; Ding, G.; Li, Y.; Qu, J.; Ma, S.; Lv, H.; Liu, Y.; Wang, W.; Dai, J.; Tang, Y.; Yu, S. *Org. Lett.* **2013**, *15*, 5206–5209.
- (12) Zhang, D.; Ge, H.; Xie, D.; Chen, R.; Zou, J. H.; Tao, X.; Dai, J. *Org. Lett.* **2013**, *15*, 1674–1677.